

Observation of the microstructure evolution during a mechanical assay on cardiac tissue

Jean-Marc ALLAIN

Equipe M Ξ DISIM

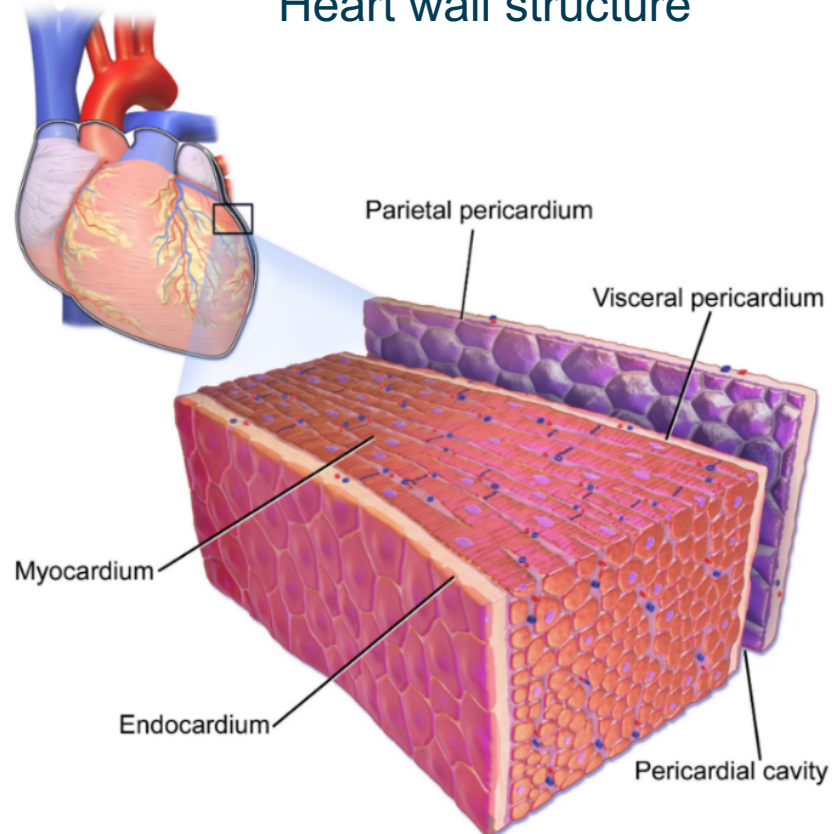
Solid Mechanics Laboratory
Ecole polytechnique, Palaiseau, France

Inria



Heart wall:

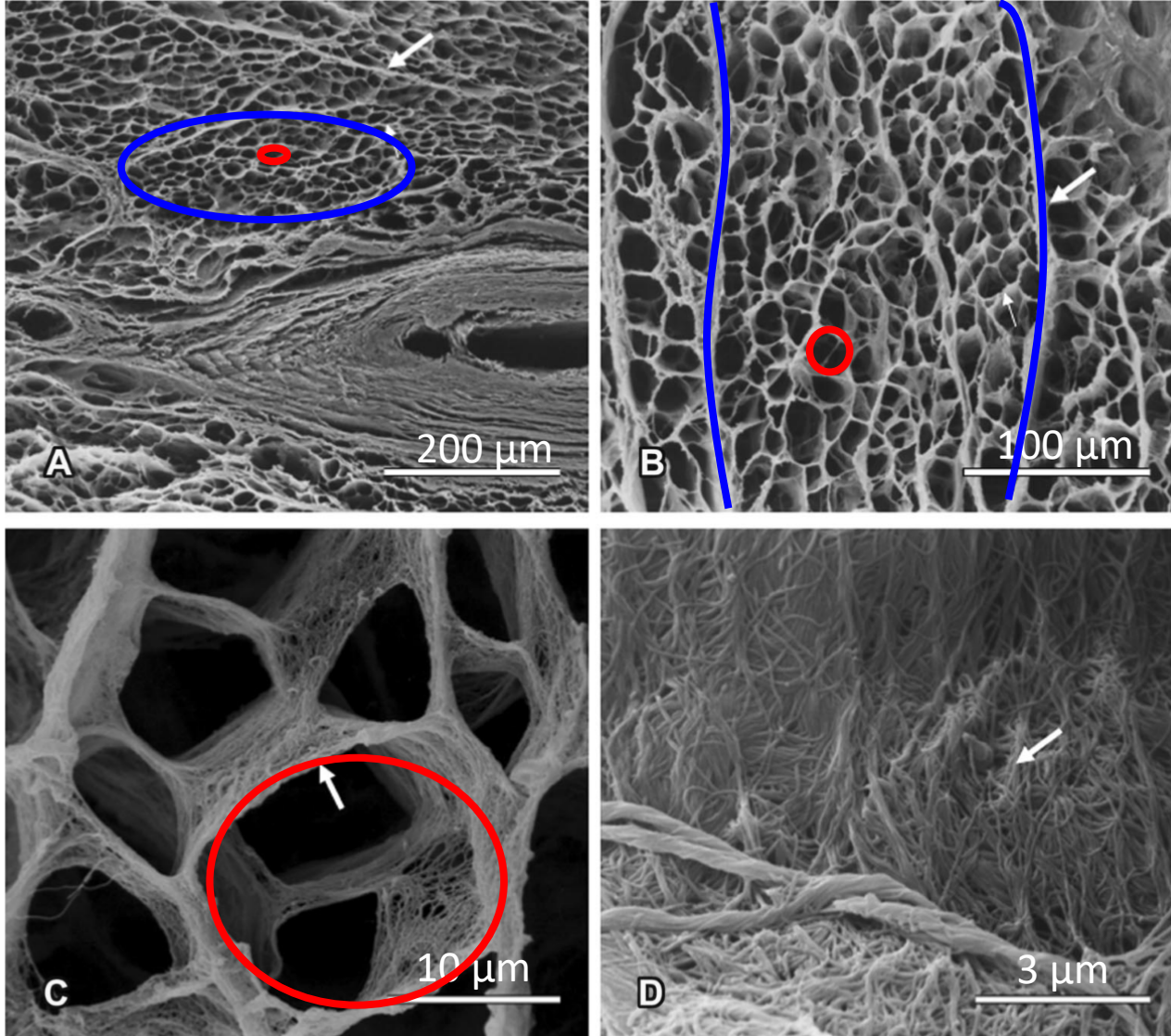
Heart wall structure



- **Cardiovascular diseases:** leading cause of death in Europe
 - **Modification of the myocardium:**
 - ↪ Structure and function closely related
 - ↪ Multiple spatial and temporal scales
- ⇒ Link between **microscopic structure** and **macroscopic mechanical properties**

Introduction

Microscopic structure: SEM observations



Fiber scale

Cardiomyocyte + ECM layer

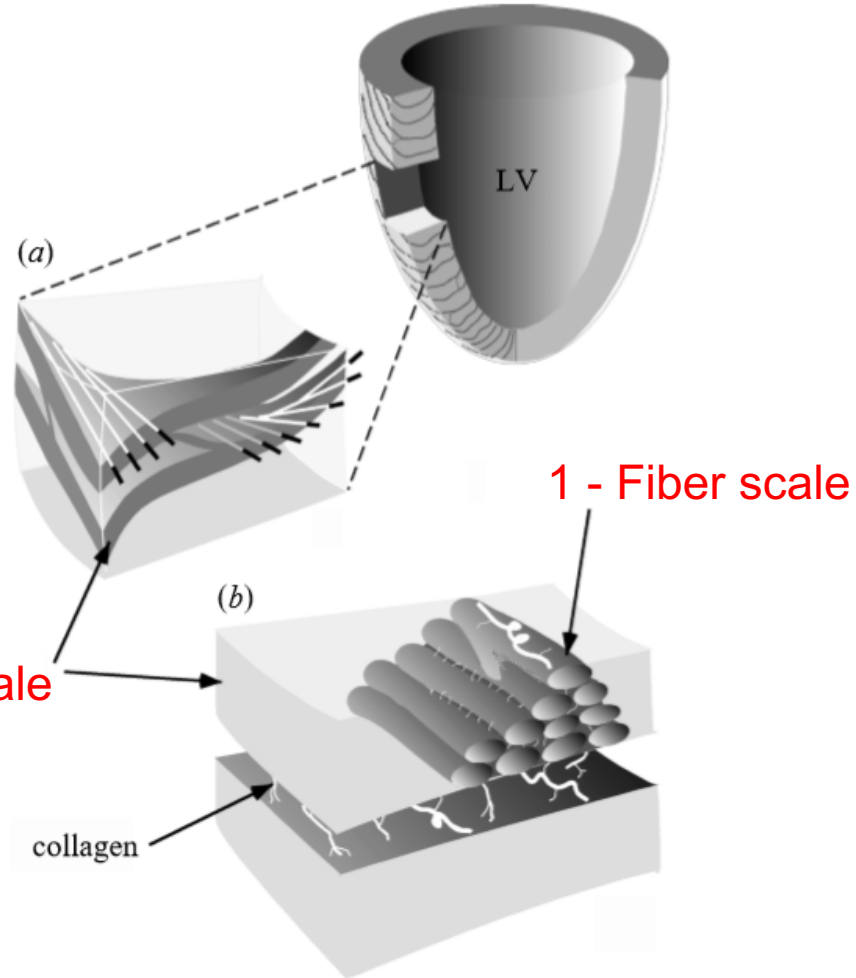
Mesosopic scale

Cardiomyocyte bundle
+ thick collagen layer

Kanzaki et al., Circulation, 2010

Introduction

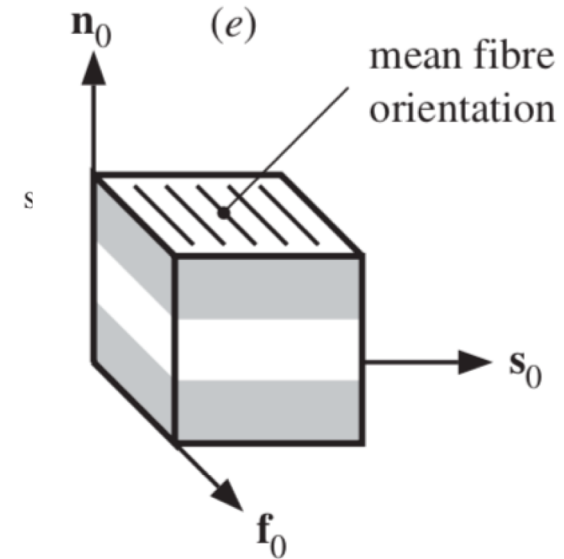
Multiscale organization



The Auckland laminar sheet model,
LeGrice et al., 1995

Ex. of model:

Holzappel & Ogden, Proc. Roy. Soc. A 2009

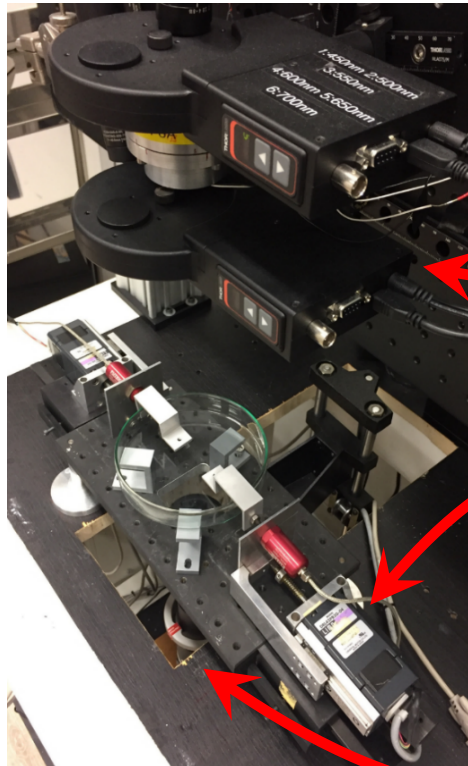


Fiber motion: **affine assumption**

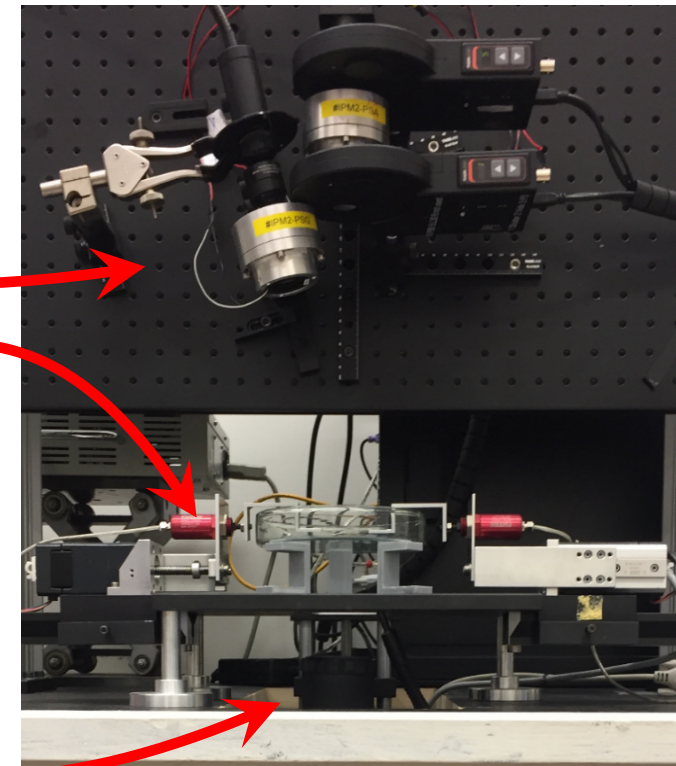
⇒ Observation of the **microstructure evolution**
during **a mechanical assay**

Simultaneous **observations** of the **mechanical** properties
and of the tissue **microstructure**

Coupled traction device with polarimetric microscope



Polarimetric setup

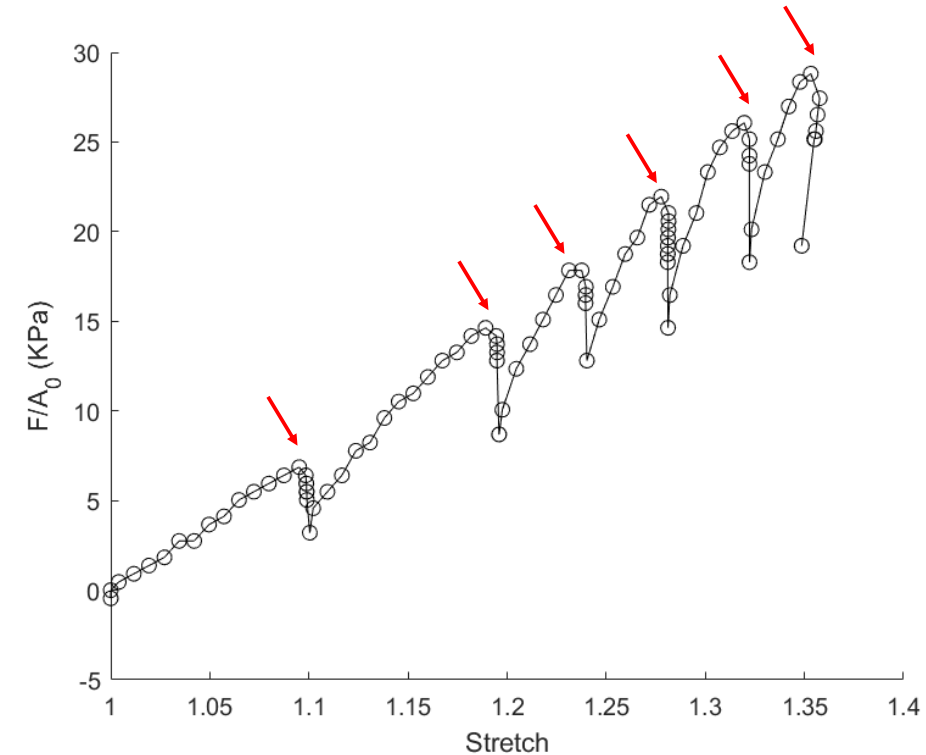


Traction device

Optical imaging

Traction

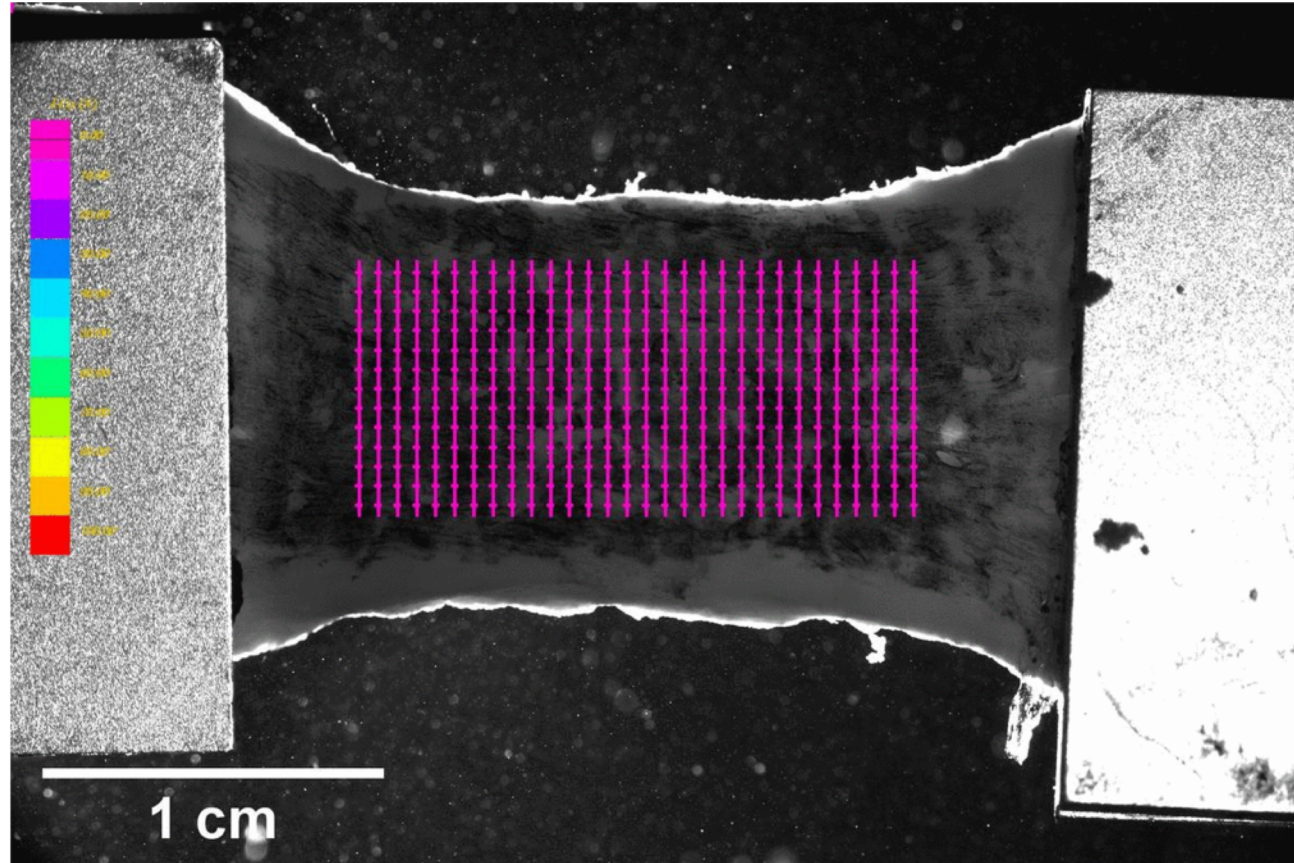
- **Sample**: pig left ventricle (30x20x2 mm)
- Sample **immersed** in PBS
- **Loading velocity**: 0.1mm/s (0.04 %/s)
- **Force** recorded every second
- **Deformation** by DIC
- **Pause** every 10% for imaging



9 samples with different initial orientations

Stretch map

- On the **lower side**
- Pixel size: $5.5 \times 5.5 \mu\text{m}^2$
- Image every 15s (0.6%)
- **ROI** ~ $16 \times 8 \text{cm}$
- **Correlation domain**: $550 \times 550 \mu\text{m}$



Maps of local deformation at different stretch levels

- ↻ **Heterogeneities**
- ↻ Stretch **comparable** on upper and lower faces

Polarimetric measure

- On the **upper side**
- Image every 10%

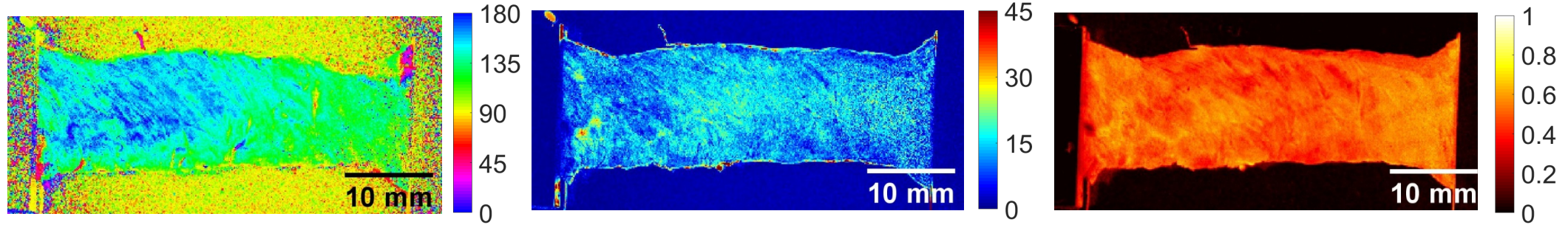
- Image: 515x385 pixels
- Pixel size: 100x100 μm^2 (sheetlet)

(a) Azimuth

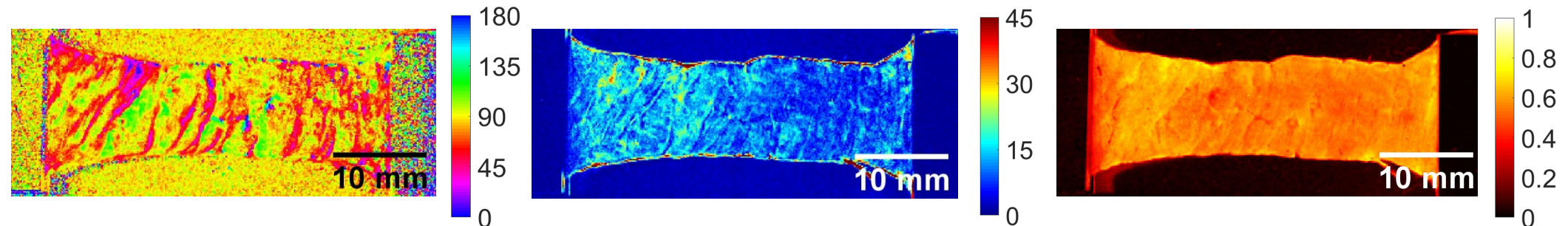
(b) Retardance

(c) Depolarization

Sample 1
 $\langle \alpha \rangle = 150^\circ$

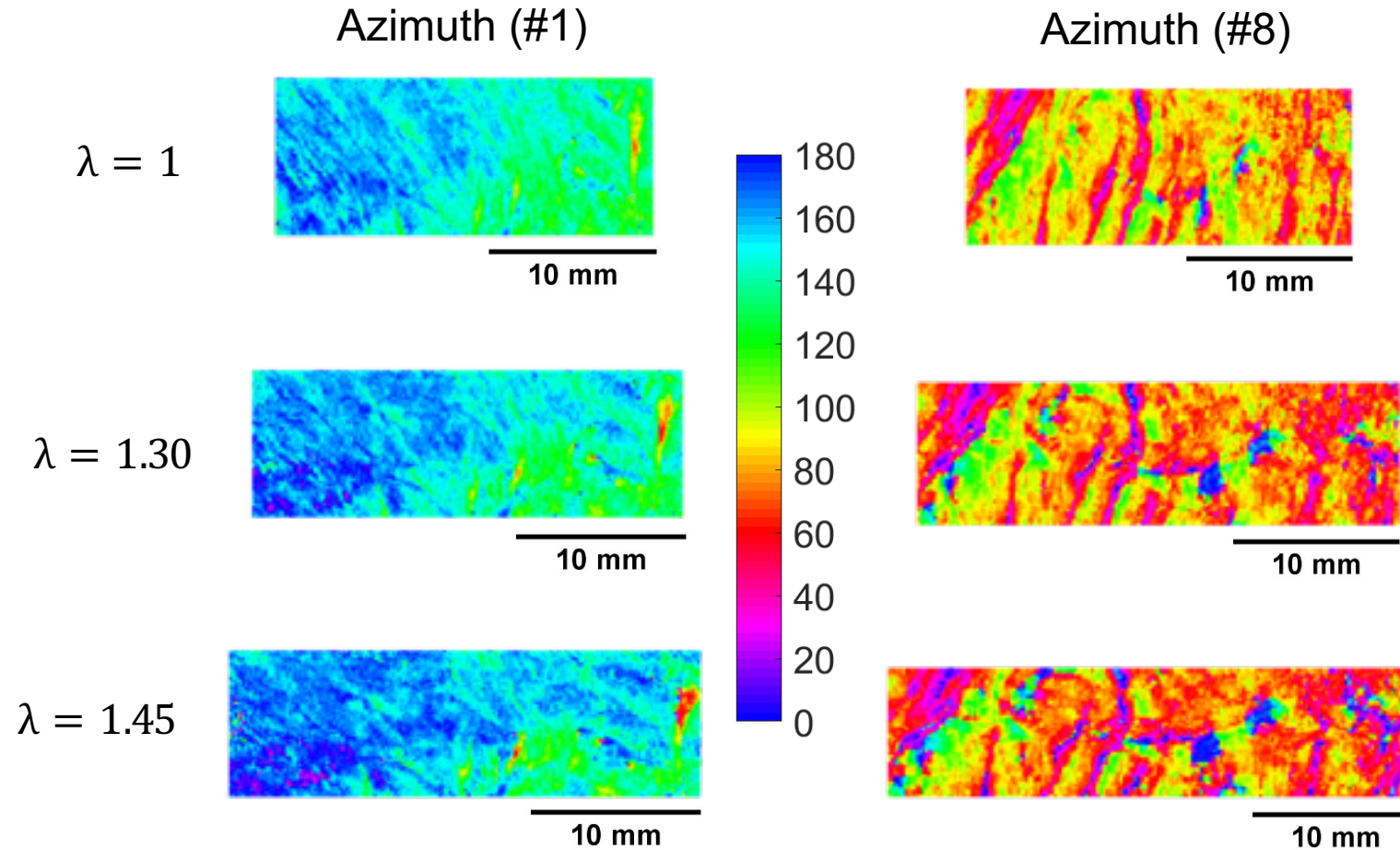


Sample 8
 $\langle \alpha \rangle = 80^\circ$



⇒ Separation lines

Evolution of the polarimetric parameters



↪ No change in retardance

↪ No change in depolarization

↪ Azimuth aligns in traction direction

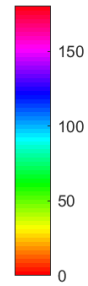
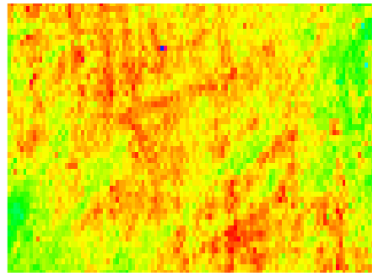
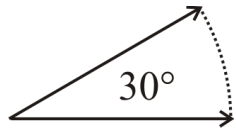
↪ Opening of the separation lines ($\langle \alpha \rangle \sim 90^\circ$)

Results

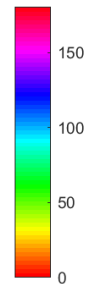
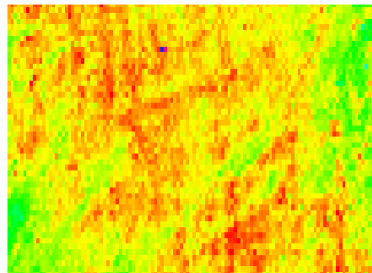
Tueni *et al.*, Sci. Report, in press

Comparison with an affine model

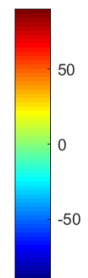
Sample #1



Measured angles (in °)

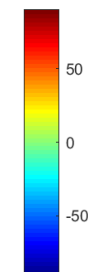
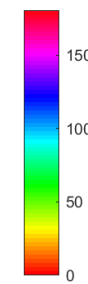
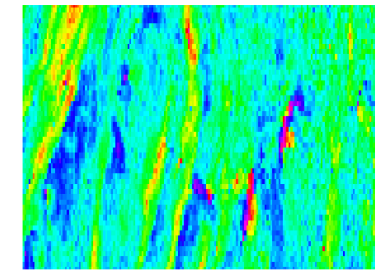
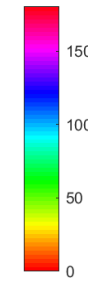
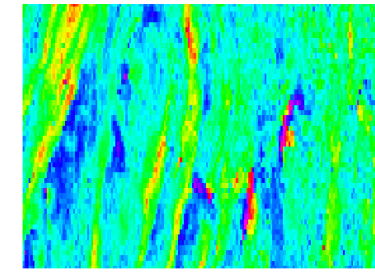
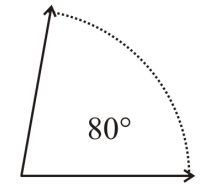


Predicted angles (in °)



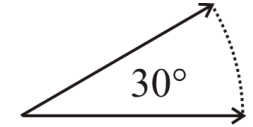
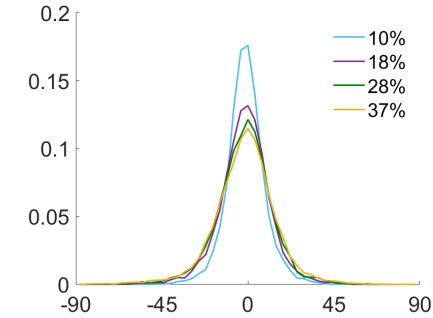
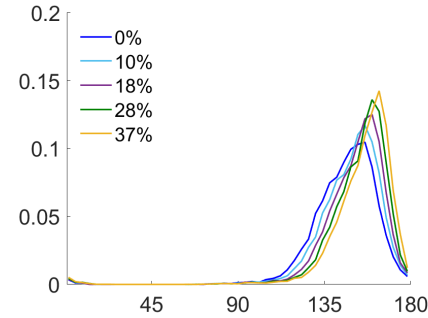
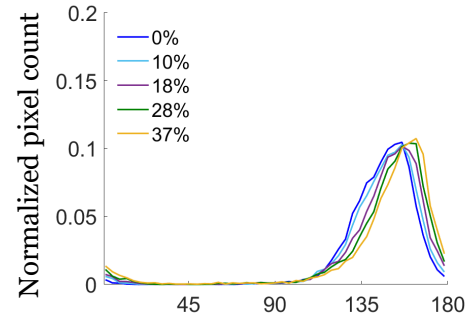
Angular difference (in °)

Sample #8

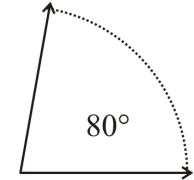
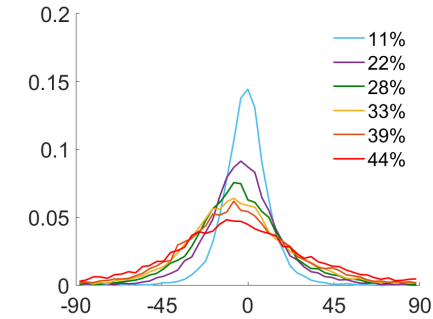
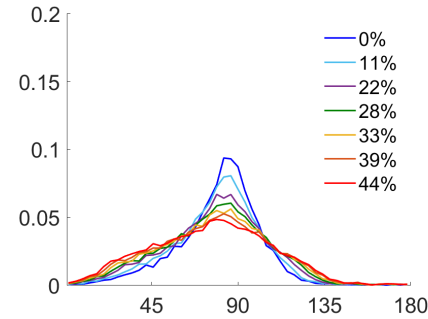
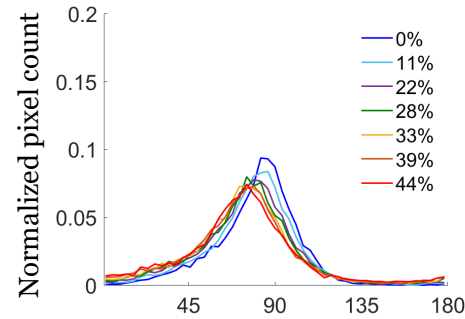


Comparison with an affine model

Sample #1

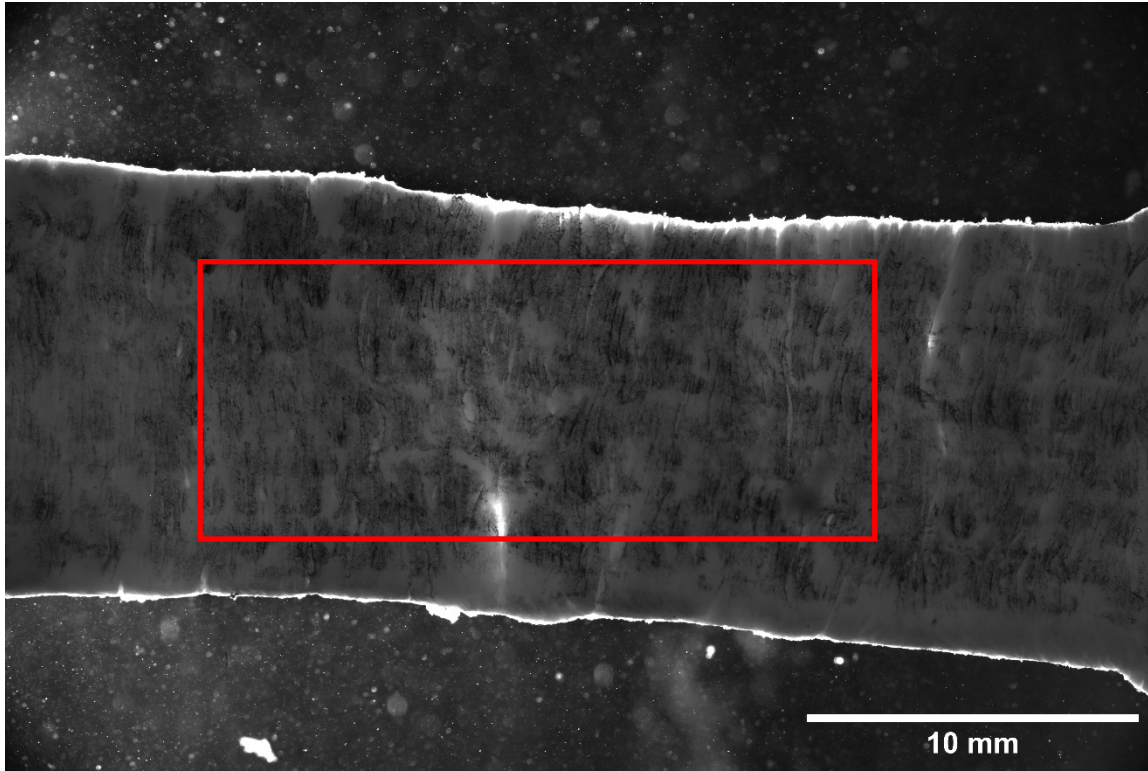


Sample #8

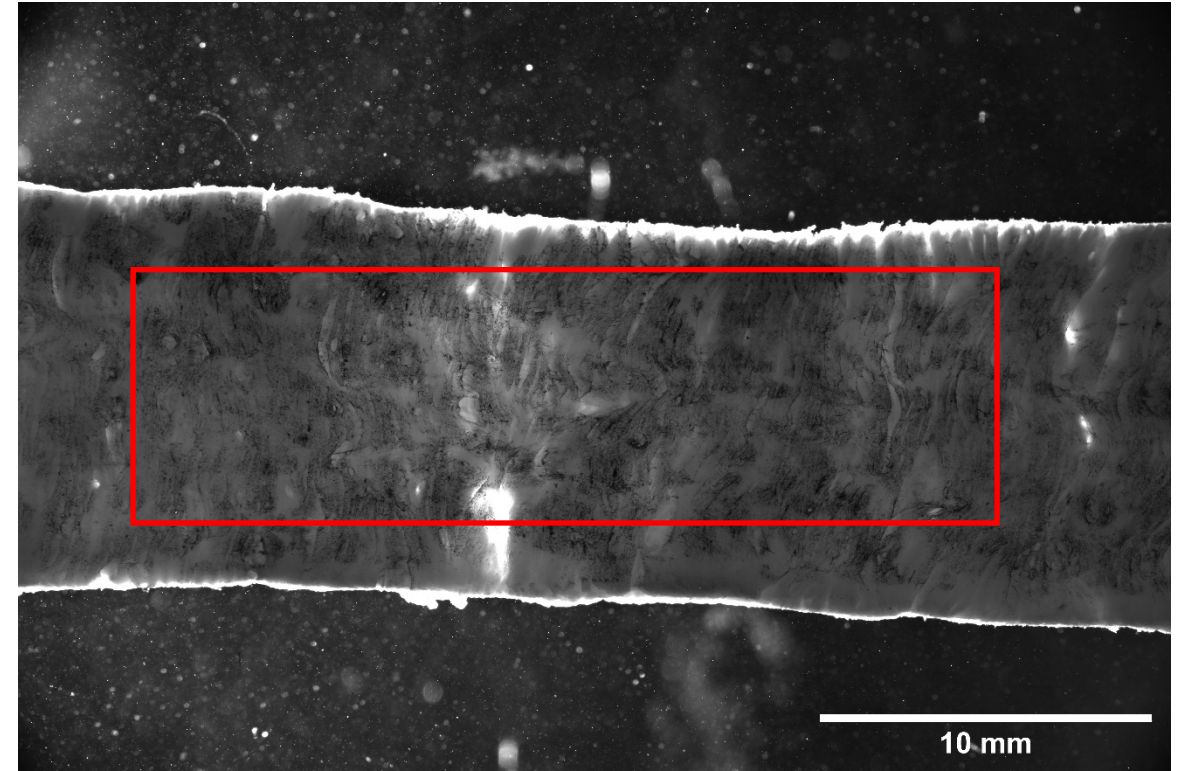


⇒ **Affine model works correctly**
apart for samples with orientation near 90°

Samples perpendicular to traction?



$\lambda = 1$



$\lambda = 1.35$

⇒ **Separation lines** open during traction

Conclusions & perspectives

➤ Conclusions

- ↪ **Multiscale observations** of myocardium
- ↪ **Affine assumption**: valid almost everywhere
- ↪ **Separation lines open under traction** (not shear or compression)

➤ Perspectives

- ↪ Other cutting **orientations** (of tissue)
- ↪ **3D organization** of the separation lines
- ↪ **Modeling** of the polarimetric signal
- ↪ Modeling of the **anisotropy** of the tissue

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